Han et al. 2025. Endemic Chinese mountain cats are threatened by domestic dogs. Cat News 84, 28-31. Supporting Online Material.

Table S1. Sample semi-structured interview questions for local herdsmen in the Gyatong Grassland

Category	Questions
Sociodemographic data	Name, Gender, Age, Education level Occupation, Income source and level Years of residence Type and number of livestock
Chinese mountain cats	Sightings of Chinese mountain cats, including geographical location and habitat characteristics, temporal occurrence (e.g., time and season of sightings), observed behavior patterns Sighting frequency and changing trend Interactions between Chinese mountain cats and domestic cats and dogs Local knowledge and beliefs about the Chinese mountain cat
Domestic dogs	Number of domestic dogs Rearing methods (e.g., chained, free-ranging) Dog abandonment Events of domestic dogs interfering with wildlife
Stray dogs	Number of stray dogs in village Population trend of stray dogs in village in recent years Potential origin of stray dogs Disease and vaccination status of stray dogs Interactions between domestic dogs and stray dogs Events of stray dogs interfering with wildlife Events of stray dogs interfering with livestock Events of stray dogs attacking people
Attitudes and management measures of stray dogs	Attitudes towards stray dogs Involvement in feeding stray dogs and (if so) its frequency Situations when stray dogs should be managed (e.g., attacking people, hunting livestock, stealing food, transmitting disease, hunting wildlife) Desired compensation for damages and losses caused by stray dogs Desired stray dog management measures (e.g., killing, neutering, shelter, adoption) Desired number of stray dogs

Table S2. PCR reaction mixture composition for the genetic analysis

Component	Volume/μL

2×EasyTaq® PCR SuperMix (TransGen Biotech, Beijing, China)	10
Forward primer [0.2 μM]	2
Reverse primer [0.2μM]	2
BSA [0.5 μg/μl]	1
Template DNA	2
Nuclease-Free Water	3
Total	20

Table S3. PCR protocol for the genetic analysis

Step	Temperature	Time	Number of cycles
Initial denaturation	95 °C	15 min	
Denaturation	95 °C	30 s	
Annealing	65-55°C	30 s	down 0.5 °C per cycle
Extension	72 °C	30 s	
Denaturation	95 °C	30 s	
Annealing	55 °C	30 s	32 cycles
Extension	72 °C	30 s	
Final extension	56 °C	30 min	

Table S4. Primers used to sequence 9 nuclear loci and partial *CytB* gene

Locus	Primer name	Sequence 5'-3'	Size /bp	Source
CLU	CLU_f	AAGGGGCTTGCTGACTGG	193	Self designed based on Johnson *et al.*, 2006
	CLU_r	AGAGCAATATAGTGATGGGCCA		
HK1	HK1_f	CGGTTGTATCCTGGTAGCCT	294	Self designed based on Johnson *et al.*, 2006
	HK1_r	CGAGCTCTCTGGTTTCATGC		
RSA2	RSA2_f	TCCGGGTTTTGTTCCATCTT	294	Self designed based on Johnson *et al.*, 2006
	RSA2_r	TGGCCATGCTAAGGGAATAAAG		
GHR	GHR_f	TTAACCTCTGTGGCTGAGCA	658	Self designed based on Johnson *et al.*, 2006
	GHR_r	TTGATCCAGATCTCCTCAAGGT		
DGKG2	DGKG2_f	GGTCGTAGTCCATTCCTTGC	700	Self designed based on Johnson *et al.*, 2006
	DGKG2_r	CAGAAGCAAGGGGTGATGTC		
GATA3	GATA3_f	TCTCTCTAGTGCTGTGAAAACAAA	440	Self designed based on Johnson *et al.*, 2006
	GATA3_r	CGGAAAAGGCTTGCTGAG		
GNB1	GNB1_f	TGCCAGCTGTTAGCGAGTTA	650	Self designed based on Johnson *et al.*, 2006

	GNB1_r	AGGGAGGGCGTCGGTACT		
RAG2	RAG2_f	GATTTATGTCATGTCTGTTGTTGG	430	Self designed based on Johnson *et al.*, 2006
	RAG2_r	CTCCTGGCAATACTGTGCAA		
PLP	PLP_f	TCATCAATGTGTAAGTACCTGTCC	820	Self designed based on Johnson *et al.*, 2006
	PLP_r	GGCATGGATCCTGCATTAAC		
CytB	H15149	AAACTGCAGCCCCTCAGAATGATA TTTGTCCTCA	403	Kocher *et al.*, 1989
	CanidL1	AATGACCAACATTCGAAA		Paxinos *et al.*, 1997

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